Package 'phyloregion'

July 19, 2020

Type Package

Title Biogeographic Regionalization and Macroecology

Version 1.0.4

Description Computational infrastructure for biogeography, community ecology,
and biodiversity conservation (Daru et al. 2020) <doi:10.1101/2020.02.12.945691>.
It is based on the methods described in Daru et al. (2020) <doi:10.1038/s41467-020-15921-6>.
The original conceptual work is described in Daru et al. (2017) <doi:10.1016/j.tree.2017.08.013>
on patterns and processes of biogeographical regionalization. Additionally, the package
contains fast and efficient functions to compute more standard conservation measures
such as phylogenetic diversity, phylogenetic endemism, evolutionary distinctiveness
and global endangerment, as well as compositional turnover (e.g., beta diversity).

Imports ape, phangorn, Matrix, betapart, parallel, methods, raster,
colorspace, vegan, sp, igraph, rgdal, rgeos

Suggests tinytest, knitr, rmarkdown, mapproj

VignetteBuilder knitr

URL https://github.com/darunabas/phyloregion,
https://darunabas.github.io/phyloregion/index.html

BugReports https://github.com/darunabas/phyloregion/issues

License AGPL-3

Encoding UTF-8

RoxygenNote 7.1.1

NeedsCompilation no

Depends R (>= 3.6.0)

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Repository CRAN

Date/Publication 2020-07-19 12:50:02 UTC

R **topics documented:**

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phyloregion-package *Biogeographic regionalization and spatial conservation*

Description

This document describes the `phyloregion` package for the R software. `phyloregion` is a computational infrastructure for biogeographic regionalization (the classification of geographical areas in terms of their biotas) and spatial conservation in the R scientific computing environment. Previous analyses of biogeographical regionalization were either focused on smaller datasets or slower particularly when the number of species or geographic scale is very large. With macroecological datasets of ever increasing size and complexity, `phyloregion` offers the possibility of handling and executing large scale biogeographic regionalization efficiently and with extreme speed. It also allows fast

and efficient for analysis of more standard conservation measures such as phylogenetic diversity, phylogenetic endemism, evolutionary distinctiveness and global endangerment. `phyloregion` can run on any operating system (Mac, Linux, Windows or even high performance computing cluster) with R 3.6.0 (or higher) installed.

How to cite `phyloregion`

The original implementation of phyloregion is described in:

- Daru B.H., Karunarathne, P. & Schliep, K. (2020) phyloregion: R package for biogeographic regionalization and spatial conservation. *bioRxiv* 2020.02.12.945691 doi: 10.1101/2020.02.12.945691

It is based on the method described in:

- Daru, B.H., Farooq, H., Antonelli, A. & Faurby, S. (2020) Endemism patterns are scale dependent. *Coming soon.*

The original conceptual is described in:

- Daru, B.H., Elliott, T.L., Park, D.S. & Davies, T.J. (2017) Understanding the processes underpinning patterns of phylogenetic regionalization. *Trends in Ecology and Evolution* **32**: 845-860.

Feedback

If you have any questions, suggestions or issues regarding the package, please add them to GitHub issues

Installation

`phyloregion` is an open-source and free package hosted on GitHub. You will need to install the `devtools` package. In R, type:

```
if (!requireNamespace("devtools",quietly = TRUE)) install.packages("devtools")
```

Then:

```
devtools::install_github("darunabas/phyloregion")
```

Load the phyloregion package:

```
library(phyloregion)
```

Acknowledgments

Barnabas Daru thanks Texas A&M University-Corpus Christi for financial and logistic support.

Author(s)

Barnabas H. Daru, Piyal Karunarathne, Klaus Schliep

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africa   *Plants of southern Africa*

Description

This dataset consists of a dated phylogeny of the woody plant species of southern Africa along with their geographical distributions. The dataset comes from a study that maps tree diversity hotspots in southern Africa (Daru et al. 2015). The study mapped five types of diversity hotspots including species richness (SR), phylogenetic diversity (PD), phylogenetic endemism (PE), species weighted endemism (CWE), and evolutionary distinctiveness and global endangerment (EDGE). The results revealed large spatial incongruence between biodiversity indices, resulting in unequal representation of PD, SR, PE, CWE and EDGE in hotspots and currently protected areas, suggesting that an integrative approach which considers multiple facets of biodiversity is needed to maximise the conservation of tree diversity in southern Africa. Specifically for this package, we arranged the dataset into four components: "comm", "polys", "phylo", "mat", "IUCN".

Details

- comm: This a sparse community composition matrix of each species presences/absences within 50 × 50 km grid cells. A sparse matrix is a matrix with a high proportion of zero entries (Duff 1977), of which only the non-zero entries are stored and used for downstream analysis.

- polys: These are the grid cells covering the geographic extent of study area. These can be created using the function `fishnet`. The polys object is of class SpatialPolygonsDataFrame and has a column labeled "grids", with the grid identities.

- phylo: This corresponds to the phylogenetic tree which was estimated using Bayesian analysis of 1,400 species and 1,633 bp of chloroplast DNA sequences derived from a combination of *matK* and *rbcLa*, assuming an uncorrelated relaxed molecular clock model, using the program BEAST v.1.7.5 (Drummond & Rambaut, 2007). Branch lengths were calibrated in millions of years using a Bayesian MCMC approach by enforcing topological constraints assuming APG III backbone from Phylomatic v.3 (Webb & Donoghue, 2005) and 18 fossil calibration points from Bell et al. (2010).

- mat: This is a distance matrix of phylogenetic beta diversity between all grid cells at the 50 × 50 km scale.

- IUCN: This is a dataframe of IUCN conservation status of each woody species (LC, NT, VU, EN, CR). This is useful for analysis of Evolutionary Distinctiveness and Global Endangerment using the function `EDGE`.

References

Bell, C.D., Soltis, D.E., & Soltis, P.S. (2010). The age and diversification of the angiosperms re-revisited. *American Journal of Botany* **97**, 1296–1303.

Daru, B.H., Van der Bank, M. & Davies, T.J. (2015) Spatial incongruence among hotspots and complementary areas of tree diversity in southern Africa. *Diversity and Distributions* **21**, 769-780.

Drummond, A.J., & Rambaut, A. (2007). BEAST: Bayesian evolutionary analysis by sampling trees. *BMC Evolutionary Biology* **7**, 214.

Duff, I.S. (1977). A survey of sparse matrix research. *Proceedings of the IEEE* **65**, 500–535.

Webb, C.O., & Donoghue, M.J. (2005). Phylomatic: Tree assembly for applied phylogenetics. *Molecular Ecology Notes* **5**, 181–183.

Examples

```
data(africa)
names(africa)

library(raster)
library(ape)
plot(africa$polys)
plot(africa$phylo)
```

---

beta_core   *Taxonomic (non-phylogenetic) beta diversity*

Description

Data are assumed to be presence / absence (0 / 1) and all values greater zero are assumed to reflect presence.

Usage

```
beta_core(x)

beta_diss(x, index.family = "sorensen")
```

Arguments

x   an object of class Matrix, where rows are sites and columns are species.

index.family   family of dissimilarity indices, partial match of "sorensen" or "jaccard".

Details

`beta_core` is helper function to compute the basic quantities needed for computing the "sorensen" or "jaccard" index.

Value

`beta_core` returns an object of class `beta_diss` like the `betapart.core` function. This object can be called by `beta.pair` or `beta.multi`.

`beta_diss` returns a list with three dissimilarity matrices. See `beta.pair` for details.

Author(s)

Klaus Schliep

See Also

[betapart.core](), [betapart](), [phylobeta]()

Examples

```
data(africa)
x <- africa$comm
bc <- beta_core(x)
beta_sorensen <- beta_diss(x)
```

---

choropleth   *Bin values*

Description

choropleth discretizes the values of a quantity for mapping.

Usage

```
choropleth(x, k = 10, breaks = ”quantile”, min = NULL, max = NULL)
```

Arguments

x	Vector of values to discretize.
k	Numeric, the desired number of bins to discretize.
breaks	one of "equal", "pretty", "jenks", "quantile" or numeric vector with the actual breaks by specifying the minimum (min) and maximum (max) bounds.
min	the minima of the lowest bound of the break.
max	the maxima of the upper bound of the break

Value

a vector with the discretized values.

Author(s)

Barnabas H. Daru <darunabas@gmail.com>

See Also

[coldspots]()

Examples

```
library(sp)
s <- readRDS(system.file("nigeria/SR_Naija.rds", package = "phyloregion"))
k <- 10
COLOUR <- hcl.colors(k, "RdYlBu")
y <- choropleth(s$SR, k)

plot(s$SR, y)
## To plot and color according to some metric:
plot(s, col = COLOUR[y])
```

---

coldspots	*Computes biodiversity coldspots and hotspots*

Description

`coldspots` and `hotspots` map areas or grid cells with lowest or highest values, respectively, of a biodiversity metric e.g. species richness, species endemism or degree of threat.

Usage

```
coldspots(x, prob = 2.5, ...)

hotspots(x, prob = 2.5, ...)
```

Arguments

x	a vector on which to compute coldspots
prob	The threshold quantile for representing the lowest (`coldspots`) or highest (`hotspots`) proportion of biodiversity in an area. By default, the threshold is set to `prob = 2.5` percent.
...	Further arguments passed to or from other methods.

Value

A vector of integers of 1s and 0s with 1 corresponding to the coldspots or hotspots

Author(s)

Barnabas H. Daru <darunabas@gmail.com>

References

Myers, M., Mittermeier, R.A., Mittermeier, C.G., da Fonseca, G.A.B. & Kent, J. (2000) Biodiversity hotspots for conservation priorities. *Nature* **403**: 853–858.

Ceballos, G. & Ehrlich, P.R. (2006) Global mammal distributions, biodiversity hotspots, and conservation. *Proceedings of the National Academy of Sciences USA* **103**: 19374–19379.

Orme, C.D., Davies, R.G., Burgess, M., Eigenbrod, F., Pickup, N. et al. (2005) Global hotspots of species richness are not congruent with endemism or threat. *Nature* **436**: 1016–1019.

Daru, B.H., Van der Bank, M. & Davies, T.J. (2015) Spatial incongruence among hotspots and complementary areas of tree diversity in southern Africa. *Diversity and Distributions* **21**: 769-780.

See Also

choropleth

Examples

```
library(raster)
library(sp)
data(africa)
names(africa)

Endm <- weighted_endemism(africa$comm)
C <- coldspots(Endm) # coldspots
H <- hotspots(Endm) # hotspots

## Merge endemism values to shapefile of grid cells.
DF <- data.frame(grids=names(C), cold=C, hot=H)
m <- merge(africa$polys, DF, by = "grids", all = TRUE)

plot(africa$polys, border = "grey", col = "lightgrey",
  main = "Weighted Endemism Hotspots and Coldspots")
plot(m[(m@data$cold == 1), ], col = "blue", add = TRUE, border = NA)
plot(m[(m@data$hot == 1), ], col = "red", add = TRUE, border = NA)
legend("bottomleft", fill = c("blue", "red", "yellow", "green"),
  legend = c("coldspots", "hotspots"), bty = "n", inset = .092)
```

---

collapse_range   *Collapse nodes and ranges based on divergence times*

Description

This function collapses nodes and geographic ranges based on species' divergence times at various time depths.

Usage

```
collapse_range(
  x,
  tree,
  n,
  species = "species",
  grids = "grids",
  format = "wide"
)
```

Arguments

x	A community matrix or data frame.
tree	A phylogenetic tree.
n	Time depth to slice the phylogenetic tree (often in millions of years for dated trees).
species	If format = "long" (the default), the column with the species name.
grids	The column with the sites or grids if format = "long".
format	Format of the community composition data: "long" or "wide" with species as columns and sites as rows.

Value

Two community data frames: the `collapsed community data` and `original community data`

References

Daru, B.H., Farooq, H., Antonelli, A. & Faurby, S. Endemism patterns are scale dependent.

Examples

```
library(ape)
tr1 <- read.tree(text ="(((a:2,(b:1,c:1):1):1,d:3):1,e:4);")
com <- matrix(c(1,0,1,1,0,0,
                1,0,0,1,1,0,
                1,1,1,1,1,1,
                1,0,1,1,0,1,
                0,0,0,1,1,0), 6, 5,
              dimnames=list(paste0("g",1:6), tr1$tip.label))

collapse_range(com, tr1, n=1)
```

EDGE   *Evolutionary Distinctiveness and Global Endangerment*

Description

This function calculates EDGE by combining evolutionary distinctiveness (ED; i.e., phylogenetic isolation of a species) with global endangerment (GE) status as defined by the International Union for Conservation of Nature (IUCN).

Usage

```
EDGE(x, phy, Redlist = "Redlist", species = "species", ...)
```

Arguments

x	a data.frame
phy	a phylogenetic tree (object of class phylo).
Redlist	column in the data frame with the IUCN ranks: LC, NT, VU, EN, CR, and EX.
species	data frame column specifying the taxon
...	Further arguments passed to or from other methods.

Details

EDGE is calculated as:

$$log(1 + ED) + GE * log(2)$$

where *ED* represents the evolutionary distinctiveness score of each species (function `evol_distinct`), i.e. the degree of phylogenetic isolation, and combining it with *GE*, global endangerment from IUCN conservation threat categories. *GE* is calculated as the expected probability of extinction over 100 years of each taxon in the phylogeny (Redding & Mooers, 2006), scaled as follows: least concern = 0.001, near threatened and conservation dependent = 0.01, vulnerable = 0.1, endangered = 0.67, and critically endangered = 0.999.

Value

Returns a dataframe of EDGE scores

Author(s)

Barnabas H. Daru

References

Redding, D.W., & Mooers, A.Ø. (2006) Incorporating evolutionary measures into conservation prioritization. *Conservation Biology* **20**: 1670–1678.

Isaac, N.J., Turvey, S.T., Collen, B., Waterman, C. & Baillie, J.E. (2007) Mammals on the EDGE: conservation priorities based on threat and phylogeny. *PLoS ONE* **2**: e296.

Examples

```
data(africa)
y <- EDGE(africa$IUCN, africa$phylo, Redlist = "IUCN", species="Species")
```

---

evol_distinct	*Species' evolutionary distinctiveness*

Description

Calculates evolutionary distinctiveness measures for a suite of species by: a) equal splits (Redding and Mooers 2006) b) fair proportions (Isaac et al., 2007). This a new implementation of the picante function `evol.distinct` however allowing multifurcations and can be orders of magnitude faster.

Usage

```
evol_distinct(
  tree,
  type = c("equal.splits", "fair.proportion"),
  scale = FALSE,
  use.branch.lengths = TRUE,
  ...
)
```

Arguments

tree	an object of class phylo.
type	a) equal splits (Redding and Mooers 2006) or b) fair proportions (Isaac et al., 2007)
scale	The scale option refers to whether or not the phylogeny should be scaled to a depth of 1 or, in the case of an ultrametric tree, scaled such that branch lengths are relative.
use.branch.lengths	
	If use.branch.lengths=FALSE, then all branch lengths are changed to 1.
...	Further arguments passed to or from other methods.

Value

a named vector with species scores.

Author(s)

Klaus Schliep

References

Redding, D.W. and Mooers, A.O. (2006). Incorporating evolutionary measures into conservation prioritisation. *Conservation Biology*, **20**, 1670–1678.

Isaac, N.J.B., Turvey, S.T., Collen, B., Waterman, C. and Baillie, J.E.M. (2007). Mammals on the EDGE: conservation priorities based on threat and phylogeny. *PLoS ONE*, **2**, e296.

See Also

[evol.distinct](), [phyloregion]()

Examples

```
tree <- ape::rcoal(10)
evol_distinct(tree)
evol_distinct(tree, type = "fair.proportion")
```

---

fishnet	*Create a fishnet of regular grids*

Description

The `fishnet` function creates a regular grid of locations covering the study area at various grain sizes.

Usage

```
fishnet(mask, res = 0.5)
```

Arguments

mask	a polygon shapefile covering the boundary of the survey region.
res	the grain size of the grid cells in decimal degrees (default).

Value

A spatial polygon object of equal area grid cells covering the defined area.

References

Phillips, S.J., Anderson, R.P. & Schapire, R.E. (2006) Maximum entropy modeling of species geographic distributions. *Ecological Modelling* **190**: 231-259.

Examples

```
file <- system.file("nigeria/nigeria.rds", package="phyloregion")
d <- readRDS(file)
d1 <- fishnet(d, res = 0.75)
```

---

get_clades   *Get descendant nodes of phylogeny at a given time depth*

Description

`get_clades` returns the tips that descend from a given node or time depth on a dated phylogenetic tree.

Usage

```
get_clades(tree, cut = NULL, k = NULL)
```

Arguments

tree	is a dated phylogenetic tree with branch lengths stored as a phylo object (as in the ape package).
cut	the slice time
k	number of slices

Value

A list of descendants

References

Schliep, K.P. (2010) phangorn: phylogenetic analysis in R. *Bioinformatics* **27**: 592–593.

Examples

```
require(ape)
data(bird.orders)
plot(bird.orders)
axisPhylo(side = 1)
abline(v=28-23) # the root is here at 28
get_clades(bird.orders, 23)
```

---

hexcols   *Generate diverging colors in HCL colour space.*

Description

A function to generate colors in Hue-Chroma-Luminance colour scheme for mapping phyloregions.

Usage

```
hexcols(x)
```

Arguments

x   An object of class [metaMDS](#)

Value

A range of discrete colors differentiating between phyloregions in terms of their shared relation-
ships.

Author(s)

Barnabas H. Daru <darunabas@gmail.com>

Examples

```
library(vegan)
data(dune)
c1 <- metaMDS(dune, trace = 0)
hexcols(c1)
plot(c1$points, pch = 21, cex = 7, bg = hexcols(c1), las = 1)
```

---

indicators   *Top driving species in phyloregions*

Description

This function applies a KL-divergence approach to a list of indicator species in phyloregions.

Usage

```
indicators(
  theta,
  top_indicators = 5,
  method = c("poisson", "bernoulli"),
  options = c("min", "max"),
  shared = FALSE
)
```

Arguments

theta	A matrix or data.frame of cluster probability distributions from a topics model-ing.
top_indicators	Integer to obtain the top driving species in clusters.
method	The model assumption for KL divergence measurement. Available choices are "poisson" (default) and "bernoulli".
options	Option "min" selects species that maximize the minimum KL divergence of a phyloregion vs all other phyloregions. Option "max" selects species that max-imize the maximum KL divergence of a phyloregion against all other phylore-gions.
shared	Logical if TRUE, lists top species driving patterns in more than one phyloregion.

Value

A list of top indicator species and their indicator values

Examples

```
data(africa)
indsp <- indicators(africa$theta, top_indicators = 5,
                    options = "max", method = "poisson")
```

---

long2sparse *Conversion of community data*

Description

These functions convert a community data to compressed sparse matrix, dense matrix and long format (e.g. species records).

Usage

```
long2sparse(x, grids = "grids", species = "species")

sparse2long(x)

dense2sparse(x)

sparse2dense(x)

long2dense(x)

dense2long(x)
```

Arguments

x	A community data which one wants to transform
grids	column name of the column containing grid cells
species	column name of the column containing the species / taxa names

Value

A compressed sparse community matrix of sites by species

Examples

```
data(africa)
africa$comm[1:5, 1:20]
long <- sparse2long(africa$comm)
long[1:5, ]
sparse <- long2sparse(long)
all.equal(africa$comm, sparse)

dense_comm <- matrix(c(1,0,1,1,0,0,
                       1,0,0,1,1,0,
                       1,1,1,1,1,1,
                       0,0,1,1,0,1), 6, 4,
                   dimnames=list(paste0("g",1:6), paste0("sp", 1:4)))
dense_comm
sparse_comm <- dense2sparse(dense_comm)
sparse_comm
sparse2long(sparse_comm)
```

---

map_trait   *Map species' trait values in geographic space*

Description

`map_trait` add species trait values to species distribution in geographic space.

Usage

```
map_trait(x, trait, FUN = sum, shp = NULL, ...)
```

Arguments

x	A community data object - a vector (with names matching trait data) or a data.frame or matrix (with column names matching names in trait data)
trait	A data.frame of species traits with a column of species names matching species names in the community data, and another column with the trait values.
FUN	The function used to aggregate species trait values in geographic space. By default, if FUN = sum, the sum of all species traits per area or grid cell is calculated.
shp	a polygon shapefile of grid cells.
...	Further arguments passed to or from other methods.

Value

A data frame of species traits by site.

Author(s)

Barnabas H. Daru <darunabas@gmail.com>

Examples

```
data(africa)
x <- EDGE(africa$IUCN, africa$phylo, Redlist = "IUCN", species="Species")
y <- map_trait(africa$comm, x, FUN = sd, shp=africa$polys)

plot_swatch(y, y$traits, k=20)
```

---

match_phylo_comm *Match taxa and in phylogeny and community matrix*

Description

match_phylo_comm compares taxa (species, labels, tips) present in a phylogeny with a community matrix. Pruning, sorting and trying to add missing species on genus level if possible to match in subsequent analysis.

Usage

```
match_phylo_comm(phy, comm, delete_empty_rows = TRUE)
```

Arguments

phy	A phylogeny
comm	A (sparse) community data matrix
delete_empty_rows	
	delete rows with no observation

Details

Based on the function of the same name in picante but allows sparse matrices and with taxa addition.

Value

A list containing the following elements, pruned and sorted to match one another:

phy	A phylogeny object of class phylo
comm	A (sparse) community data matrix

Examples

```
data(africa)
tree <- africa$phylo
x <- africa$comm

subphy <- match_phylo_comm(tree, x)$phy
submat <- match_phylo_comm(tree, x)$com
```

---

mean_dist   *Mean distance matrix from a set of distance matrices*

Description

This function generates the mean pairwise distance matrix from a set many pairwise distance matrices. Note: all matrices should be of the same dimension.

Usage

```
mean_dist(files, tips, trace = 1, ...)
```

Arguments

files	list of pairwise distance matrices stored as CSVs or .rds with the same dimensions.
tips	list of site or grid names
trace	Trace the function; trace = 2 or higher will be more voluminous.
...	Further arguments passed to or from other methods.

Value

average distance matrix

---

optimal_phyloregion   *Determine optimal number of clusters*

Description

This function divides the hierarchical dendrogram into meaningful clusters ("phyloregions"), based on the 'elbow' or 'knee' of an evaluation graph that corresponds to the point of optimal curvature.

Usage

```
optimal_phyloregion(x, method = "average", k = 20)
```

Arguments

x	a numeric matrix, data frame or "dist" object.
method	the agglomeration method to be used. This should be (an unambiguous abbreviation of) one of "ward.D", "ward.D2", "single", "complete", "average" (= UPGMA), "mcquitty" (= WPGMA), "median" (= WPGMC) or "centroid" (= UPGMC).
k	numeric, the upper bound of the number of clusters to compute. DEFAULT: 20 or the number of observations (if less than 20).

Value

a list containing the following as returned from the GMD package (Zhao et al. 2011):

- k: optimal number of clusters (bioregions)
- totbss: total between-cluster sum-of-square
- tss: total sum of squares of the data
- ev: explained variance given k

References

Salvador, S. & Chan, P. (2004) *Determining the number of clusters/segments in hierarchical clustering/segmentation algorithms*. Proceedings of the Sixteenth IEEE International Conference on Tools with Artificial Intelligence, pp. 576–584. Institute of Electrical and Electronics Engineers, Piscataway, New Jersey, USA.

Zhao, X., Valen, E., Parker, B.J. & Sandelin, A. (2011) Systematic clustering of transcription start site landscapes. *PLoS ONE* **6**: e23409.

Examples

```
data(africa)
tree <- africa$phylo
bc <- beta_diss(africa$comm)
(d <- optimal_phyloregion(bc[[1]]))
plot(d$df$k, d$df$ev, ylab = "Explained variances",
  xlab = "Number of clusters")
lines(d$df$k[order(d$df$k)], d$df$ev[order(d$df$k)], pch = 1)
points(d$optimal$k, d$optimal$ev, pch = 21, bg = "red", cex = 3)
points(d$optimal$k, d$optimal$ev, pch = 21, bg = "red", type = "h")
```

PD   *Phylogenetic diversity*

Description

PD calculates Faith's (1992) phylogenetic diversity.

Usage

```
PD(x, phy)
```

Arguments

x	a community matrix, i.e. an object of class matrix or Matrix.
phy	a phylogenetic tree (object of class phylo).

Value

a vector with the PD for all samples.

References

Faith, D.P. (1992) Conservation evaluation and phylogenetic diversity. *Biological Conservation* **61**: 1–10.

See Also

read.community read.tree phylobeta_core

Examples

```
library(ape)
library(Matrix)
tree <- read.tree(text ="((t1:1,t2:1)N2:1,(t3:1,t4:1)N3:1)N1;")
com <- sparseMatrix(c(1,3,4,1,4,5,1,2,3,4,5,6,3,4,6),
  c(1,1,1,2,2,2,3,3,3,3,3,3,4,4,4),x=1,
  dimnames = list(paste0("g", 1:6), tree$tip.label))

PD(com, tree)
```

---

phylobeta_core   *Phylogenetic beta diversity*

Description

phylobeta_core computes efficiently for large community matrices and trees the necessary quantities used by the betapart package to compute pairwise and multiple-site phylogenetic dissimilarities.

Usage

```
phylobeta_core(x, phy)

phylobeta(x, phy, index.family = "sorensen")
```

Arguments

x	an object of class Matrix or matrix
phy	a phylogenetic tree (object of class phylo)
index.family	family of dissimilarity indices, partial match of "sorensen" or "jaccard".

Value

phylobeta_core returns an object of class "phylo.betapart", see phylo.betapart.core for details. This object can be called by phylo.beta.pair or phylo.beta.multi.

phylobeta returns a list with three phylogenetic dissimilarity matrices. See phylo.beta.pair for details.

Author(s)

Klaus Schliep

See Also

[read.community](#), [phylo.betapart.core](#), [beta_core](#)

Examples

```
library(ape)
tree <- read.tree(text = "((t1:1,t2:1)N2:1,(t3:1,t4:1)N3:1)N1;")
com <- sparseMatrix(c(1,3,4,1,4,5,1,2,3,4,5,6,3,4,6),
  c(1,1,1,2,2,2,3,3,3,3,3,3,4,4,4),x=1,
  dimnames = list(paste0("g", 1:6), tree$tip.label))
com

pbc <- phylobeta_core(com, tree)
pb <- phylobeta(com, tree)
```

---

phylobuilder *Create a subtree with largest overlap from a species list.*

Description

phylobuilder creates a subtree with largest overlap from a species list. If species in the species list
are not already in the tip label, species will be added at the most recent common ancestor at the
genus or family level when possible.

Usage

```
phylobuilder(species, tree, extract = TRUE)
```

Arguments

species	A vector or matrix containing a species list
tree	a phylogenetic tree (object of class phylo)
extract	extract the species in the list after trying to add missing labels to the tree. If FALSE phylobuilder adds only the taxa in the list.

Value

phylobuilder returns a phylogenetic tree, i.e. an object of class phylo.

See Also

[add.tips](#), [label2table](#), [stripLabel](#)

Examples

```
library(ape)
txt <- "(((((Panthera_leo,Panthera_pardus), Panthera_onca),(Panthera_uncia,
  (Panthera_tigris_altaica, Panthera_tigris_amoyensis)))Panthera)Felidae,
  (((((((Canis_lupus,Canis_lupus_familiaris),Canis_latrans),Canis_anthus),
  Canis_aureus),Lycaon_pictus),(Canis_adustus,Canis_mesomelas))Canis)
  Canidae)Carnivora;"
txt <- gsub("[[:space:]]", "", txt)
cats_and_dogs <- read.tree(text=txt)
plot(cats_and_dogs, node.depth=2, direction="downwards")
nodelabels(cats_and_dogs$node.label, frame="none", adj = c(0.5, 0))

tree <- drop.tip(cats_and_dogs, c("Panthera_uncia", "Lycaon_pictus"),
  collapse.singles=FALSE)

dogs <- c("Canis_lupus", "Canis_lupus_familiaris", "Canis_latrans",
  "Canis_anthus", "Canis_aureus", "Lycaon_pictus", "Canis_adustus",
  "Canis_mesomelas")

# try to extract tree with all 'dogs'
t1 <- phylobuilder(dogs, tree)
plot(t1, direction="downwards")
attr(t1, "species_list")

# providing extra information ("Family", "Order", ...) can help
sp <- data.frame(Order = c("Carnivora", "Carnivora", "Carnivora"),
  Family = c("Felidae", "Canidae", "Canidae"),
  Genus = c("Panthera", "Lycaon", "Vulpes"),
  Species = c("uncia", "pictus", "vulpes"),
  Common_name = c("Snow leopard", "Africa wild dog", "Red fox"))
sp
# Now we just add some species
t2 <- phylobuilder(sp, tree, extract=FALSE)
plot(t2, direction="downwards")
attr(t2, "species_list")
```

---

phyloregion   *Calculate evolutionary distinctiveness of phyloregions*

Description

This function estimates evolutionary distinctiveness of each phyloregion by computing the mean value of phylogenetic beta diversity between a focal phyloregion and all other phyloregions in the study area.

Usage

```
phyloregion(x, k = 10, method = "average", shp = NULL, ...)

infomap(x, shp = NULL, ...)
```

Arguments

x	A distance matrix
k	The desired number of phyloregions, often as determined by `optimal_phyloregion`.
method	the agglomeration method to be used. This should be (an unambiguous abbreviation of) one of "ward.D", "ward.D2", "single", "complete", "average" (= UPGMA), "mcquitty" (= WPGMA), "median" (= WPGMC) or "centroid" (= UPGMC).
shp	a polygon shapefile of grid cells.
...	Further arguments passed to or from other methods.

Value

An object of class `phyloregion` containing

- a data frame membership with columns grids and cluster
- k the number of clusters and additionally there can be an shape file and other bjects. This representation may still change.

Author(s)

Barnabas H. Daru <darunabas@gmail.com>

References

Daru, B.H., Van der Bank, M., Maurin, O., Yessoufou, K., Schaefer, H., Slingsby, J.A. & Davies, T.J. (2016) A novel phylogenetic regionalization of the phytogeographic zones of southern Africa reveals their hidden evolutionary affinities. *Journal of Biogeography* **43**: 155-166.

Daru, B.H., Elliott, T.L., Park, D.S. & Davies, T.J. (2017) Understanding the processes underpinning patterns of phylogenetic regionalization. *Trends in Ecology and Evolution* **32**: 845-860.

Daru, B.H., Holt, B.G., Lessard, J.P., Yessoufou, K. & Davies, T.J. (2017) Phylogenetic regionalization of marine plants reveals close evolutionary affinities among disjunct temperate assemblages. *Biological Conservation* **213**: 351-356.

See Also

evol_distinct, optimal_phyloregion, evol.distinct for a different approach.

Examples

```
library(ape)
tree <- read.tree(text = "((t1:1,t2:1)N2:1,(t3:1,t4:1)N3:1)N1;")
com <- sparseMatrix(c(1,3,4,1,4,5,1,2,3,4,5,6,3,4,6),
  c(1,1,1,2,2,2,3,3,3,3,3,3,4,4,4),x=1,
  dimnames = list(paste0("g", 1:6), tree$tip.label))
pbc <- phylobeta(com, tree)
phyloregion(pbc[[1]], k = 3)
```

---

phylo_endemism   *Phylogenetic Endemism*

Description

Calculates phylogenetic endemism (sum of 'unique' branch lengths) of multiple ecological samples.

Usage

```
phylo_endemism(x, phy, weighted = TRUE)
```

Arguments

x	is the community data given as a data.frame or matrix with species/OTUs as columns and samples/sites as rows (like in the vegan package). Columns are labeled with the names of the species/OTUs. Rows are labelled with the names of the samples/sites. Data can be either abundance or incidence (0/1). Column labels must match tip labels in the phylogenetic tree exactly!
phy	a (rooted) phylogenetic tree (phylo) with branch lengths
weighted	is a logical indicating whether weighted endemism (default) or strict endemism should be calculated.

Details

Takes a community data table and a (rooted) phylogenetic tree (with branch lengths) and calculates either strict or weighted endemism in Phylogenetic Diversity (PD). Strict endemism equates to the total amount of branch length found only in the sample/s and is described by Faith et al. (2004) as PD-endemism. Weighted endemism calculates the "spatial uniqueness" of each branch in the tree by taking the reciprocal of its range, multiplying by branch length and summing for all branch lengths present at a sample/site. Range is calculated simply as the total number of samples/sites at which the branch is present. This latter approach is described by Rosauer et al. (2009) as Phylogenetic endemism.

Value

phylo_endemism returns a vector of phylogenetic endemism for each sample or site.

References

Faith, D.P., Reid, C.A.M. & Hunter, J. (2004) Integrating phylogenetic diversity, complementarity, and endemism for conservation assessment. *Conservation Biology* **18**(1): 255-261.

Rosauer, D., Laffan, S.W., Crisp, M.D., Donnellan, C. & Cook, L.G. (2009). Phylogenetic endemism: a new approach for identifying geographical concentrations of evolutionary history. *Molecular Ecology* **18**(19): 4061-4072.

Daru, B.H., Farooq, H., Antonelli, A. & Faurby, S. (2020) Endemism patterns are scale dependent. *Nature Communications* **11** : 2115.

Examples

```
data(africa)
pe <- phylo_endemism(africa$comm, africa$phylo)
plot(density(pe))
```

---

plot.phyloregion   *Visualize biogeographic patterns*

Description

Visualize biogeographic patterns

Usage

```
## S3 method for class 'phyloregion'
plot(x, shp = NULL, palette = "NMDS", ...)

plot_NMDS(x, ...)

text_NMDS(x, ...)
```

Arguments

x	an object of class phyloregion from `phyloregion`
shp	a polygon shapefile of grid cells.
palette	name of the palette to generate colors from. The default, "NMDS", allows display of phyloregions in multidimensional scaling color space matching the color vision of the human visual system. The name is matched to the list of available color palettes from the `hcl.colors` function in the `grDevices` package.
...	arguments passed among methods.

Value

No return value, called for plotting.

Examples

```
data(africa)
tree <- africa$phylo
x <- africa$comm

subphy <- match_phylo_comm(tree, x)$phy
submat <- match_phylo_comm(tree, x)$com

pbc <- phylobeta(submat, subphy)
y <- phyloregion(pbc[[1]], shp=africa$polys)
```

```
plot_NMDS(y, cex=6)
text_NMDS(y, cex=2)
plot(y, cex=1, palette="NMDS")
plot(y, cex=1)
```

---

plot_swatch   *Plot shapefile polygons based on slot values*

Description

`plot_swatch` maps discretized values of a quantity based on their quantiles.

Usage

```
plot_swatch(
  x,
  values = NULL,
  k = 10,
  palette = "Blue-Red 3",
  key_label = "",
  leg = 10,
  lwd = 15,
  pos = "bottomleft",
  legend = TRUE,
  border = par("fg"),
  breaks = "quantile",
  min = NULL,
  max = NULL,
  ...
)
```

Arguments

x	A data frame or object of the class SpatialPolygonsDataFrame
values	Variable in the SpatialPolygonsDataFrame for which to discretize the values of the quantity.
k	Numeric, the desired number of bins to discretize.
palette	name of the palette to generate colors from. The name is matched to the list of available color palettes from the `hcl.colors` function.
key_label	label for the color key
leg	Numeric, length of the legend
lwd	numeric, line width of the legend.
pos	location to position the legend such as "bottomright", "bottomleft", "topleft", and "topright".
legend	logical indicating whether to add a legend to the map.

border	plot polygons in SpatialPolygons object
breaks	one of "equal", "pretty", "jenks", "quantile" or numeric with the actual breaks by specifying the minimum (min) and maximum (max) bounds.
min	the minima of the lowest bound of the break.
max	the maxima of the upper bound of the break
...	Further arguments passed to or from other methods.

Value

Returns no value, just map swatch of colors in geographic space!

Author(s)

Barnabas H. Daru <darunabas@gmail.com>

See Also

SpatialPolygons-class

Examples

```
library(sp)
s <- readRDS(system.file("nigeria/SR_Naija.rds", package = "phyloregion"))
plot_swatch(s, values = s$SR, k = 20)
```

---

random_species	*Generate random species distributions in space*

Description

This function generates random species distributions in geographic space as extent of occurrence range polygons based on convex hulls of random points.

Usage

```
random_species(n, species, shp, ...)
```

Arguments

n	vector of one or more elements to choose from, or a positive integer.
species	the desired number of species.
shp	the polygon shapefile of the study area for determining the species distributions
...	Further arguments passed to or from other methods.

Value

A polygon shapefile of species' extent of occurrence ranges.

Author(s)

Barnabas H. Daru <darunabas@gmail.com>

---

raster2comm   *Convert raw input distribution data to community*

Description

The functions `points2comm`, `polys2comm`, `raster2comm` provide convenient interfaces to convert raw distribution data often available as point records, polygons and raster layers, respectively, to a community composition data frame at varying spatial grains and extents for downstream analyses.

Usage

```
raster2comm(files)

polys2comm(dat, res = 1, species = "species", trace = 1, ...)

points2comm(
  dat,
  mask = NULL,
  res = 1,
  lon = "decimallongitude",
  lat = "decimallatitude",
  species = "species",
  shp.grids = NULL,
  ...
)
```

Arguments

files	list of raster layer objects with the same spatial extent and resolution.
dat	layers of merged maps corresponding to species polygons for `polys2comm`; or point occurrence data frame for `points2comm`, with at least three columns:
	• Column 1: species (listing the taxon names)
	• Column 2: decimallongitude (corresponding to decimal longitude)
	• Column 3: decimallatitude (corresponding to decimal latitude)
res	the grain size of the grid cells in decimal degrees (default).
species	a character string. The column with the species or taxon name. Default = "species".
trace	Trace the function; trace = 2 or higher will be more voluminous.
...	Further arguments passed to or from other methods.
mask	Only applicable to `points2comm`. If supplied, a polygon shapefile covering the boundary of the survey region.

lon	character with the column name of the longitude.
lat	character with the column name of the latitude.
shp.grids	if specified, the polygon shapefile of grid cells with a column labeled "grids".

Value

- comm_dat: (sparse) community matrix
- poly_shp: shapefile of grid cells with the values per cell.

See Also

mapproject for conversion of latitude and longitude into projected coordinates system. long2sparse for conversion of community data.

Examples

```
fdir <- system.file("NGAplants", package="phyloregion")
files <- file.path(fdir, dir(fdir))
ras <- raster2comm(files)
head(ras[[1]])



s <- readRDS(system.file("nigeria/nigeria.rds", package="phyloregion"))
sp <- random_species(100, species=5, shp=s)
pol <- polys2comm(dat = sp, species = "species", trace=0)
head(pol[[1]])


s <- readRDS(system.file("nigeria/nigeria.rds", package = "phyloregion"))

set.seed(1)
m <- data.frame(sp::spsample(s, 10000, type = "nonaligned"))
names(m) <- c("lon", "lat")
species <- paste0("sp", sample(1:1000))
m$taxon <- sample(species, size = nrow(m), replace = TRUE)

pt <- points2comm(dat = m, mask = s, res = 0.5, lon = "lon", lat = "lat",
            species = "taxon")
head(pt[[1]])
```

---

read.community	*Read in sparse community matrices*

Description

read.community reads in file containing occurrence data and returns a sparse matrix.

Usage

```
read.community(file, grids = "grids", species = "species", ...)
```

Arguments

file	A file name.
grids	Column name of the column containing grid cells.
species	Column name of the column containing the species / taxa names.
...	further arguments passed to or from other methods.

Value

`read.community` returns a sparse matrix (an object of class "dgCMatrix").

Examples

```
df <- data.frame(grids=paste0("g", c(1,1,2,3,3)),
                 species = paste0("sp", c(1,3,2,1,4)))
df
tmp <- tempfile()
write.csv(df, tmp)
(M <- read.community(tmp) )
sparse2long(M)
unlink(tmp)
```

---

selectbylocation   *Select features within polygon from another layer*

Description

The `selectbylocation` function selects features based on their location relative to features in another layer.

Usage

```
selectbylocation(x, y)
```

Arguments

x	source layer of the class SpatialPolygonsDataFrame or SpatialPointsDataFrame
y	Target layer or mask extent to subset from.

Value

A spatial polygons or spatial points object pruned to the extent of the target layer.

Examples

```
library(raster)
file <- system.file("nigeria/nigeria.rds", package = "phyloregion")
d <- readRDS(file)
e <- extent(d)

set.seed(1)
m <- data.frame(lon = runif(1000, e[1], e[2]),
  lat = runif(1000, e[3], e[4]),
  sites = seq(1000))
coordinates(m) <- ~ lon + lat
z <- selectbylocation(m, d)
plot(d)
points(m, col = "blue", pch = "+")
points(z, col = "red", pch = "+")
```

---

select_linkage   *Cluster algorithm selection and validation*

Description

This function contrasts different hierarchical clustering algorithms on the phylogenetic beta diversity matrix for degree of data distortion using Sokal & Rohlf's (1962) cophenetic correlation coefficient.

Usage

```
select_linkage(x)
```

Arguments

x   a numeric matrix, data frame or "dist" object.

Value

- A numeric value corresponding to the good clustering algorithm for the distance matrix

- If plot = TRUE, a barplot of cophenetic correlation for all the clustering algorithms is drawn.

References

Sokal, R.R. & Rohlf, F.J. (1962) The comparison of dendrograms by objective methods. *Taxon* **11**: 33–40.

Examples

```
data(africa)
tree <- africa$phylo
bc <- beta_diss(africa$comm)
y <- select_linkage(bc[[1]])
barplot(y, horiz = TRUE, las = 1)
```

---

timeslice   *Slice phylogenetic tree at various time depths*

Description

This function slices a dated phylogenetic tree at successive time depths back in time by collapsing younger phylogenetic branches into older ones to infer the origins of species assemblages.

Usage

```
timeslice(phy, n = 0.2, collapse = FALSE, ...)
```

Arguments

phy	A dated phylogenetic tree as an object of class "phylo".
n	Time depth to slice the phylogenetic tree (often in millions of years for dated trees).
collapse	Logical, collapse internal edges with zero edge length.
...	arguments passed among methods.

Value

A tree with the phylogenetic structure removed at the specified time depth

Author(s)

Barnabas H. Daru <darunabas@gmail.com>

References

Daru, B.H., van der Bank, M. & Davies, T.J. (2018) Unravelling the evolutionary origins of biogeographic assemblages. *Diversity and Distributions* **24**: 313–324.

Examples

```
library(ape)

set.seed(1)
tree <- rcoal(50)
x <- timeslice(tree, .5)

old.par <- par(no.readonly = TRUE)
par(mfrow = c(1, 2))
plot(tree)
axisPhylo()
plot(x)
axisPhylo()
par(old.par)
```

---

weighted_endemism   *Measure the distribution of narrow-ranged or endemic species.*

Description

weighted_endemism is species richness inversely weighted by species ranges.

Usage

```
weighted_endemism(x)
```

Arguments

x   A (sparse) community matrix.

Value

A data frame of species traits by site.

References

Crisp, M.D., Laffan, S., Linder, H.P. & Monro, A. (2001) Endemism in the Australian flora. *Journal of Biogeography* **28**: 183–198.

Daru, B.H., Farooq, H., Antonelli, A. & Faurby, S. (2020) Endemism patterns are scale dependent. *Nature Communications* **11** : 2115.

Examples

```
library(raster)
data(africa)
Endm <- weighted_endemism(africa$comm)
m <- merge(africa$polys, data.frame(grids=names(Endm), WE=Endm), by="grids")
m <- m[!is.na(m@data$WE),]
```

```
plot_swatch(m, values = m$WE, k=20)
```

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